

Electrophoretic deposition of SnO/ZnO coated Titanium for antibacterial and hemocompatible implant applications

Pragya¹, Prathiba R², Kalaiyaran M^{3*}

^{1,2,3} Department of Oral Pathology, Saveetha Dental College and Hospital, Saveetha Institute of Medical and Technical Sciences

Email: kalaiyaranm.sdc@saveetha.com

Cite this paper as: Pragya, Prathiba R, Kalaiyaran M (2024) Electrophoretic deposition of SnO/ZnO coated Titanium for antibacterial and hemocompatible implant applications. *Frontiers in Health Informatics, Vol.13, No.8*, 8088-8093

ABSTRACT

Aim: In the biomedical field, the primary objective is to develop versatile biomaterials which are capable of action against bacterial growth. Among the various challenges, chronic infections stand out as the most severe and destructive complications associated with biomaterial usage. This issue is of utmost significance in the field of orthopedics and dental implants.

Materials and methods: In this present study, we develop a biocompatible and antibacterial coating that was fabricated by electrophoretic deposition (EPD). The coating formation was investigated by surface characterization studies including surface morphology and functional group analysis which confirmed SnO/ZnO on the Cp-Ti surface.

Results: The surface exhibited a laminar array with petal-like morphology which enhances the osteointegration process. In-vitro analysis of hemocompatibility studies exhibited the coating as a hemocompatible material. Antibacterial studies showed that coating acted against *S.aureus* and *E.coli* bacteria.

Conclusion: In our study, the electrophoretic deposition of SnO/ZnO on Cp-Ti surfaces emerges as a promising biocompatible coating. This coating has demonstrated its ability to improve the surface characteristics and antibacterial efficacy of the implant.

Keywords: Titanium implant, Antibacterial, Hemocompatibility, Dental implant, Tin oxide and Zinc oxide.

INTRODUCTION

Biomaterials typically describe as materials that have been purposefully designed to control and influence the progression of diagnostic or therapeutic procedures through the regulation of their interactions with elements within living systems (Williams, 2009). Biomaterials are classified into different types such as metals, ceramics, polymers, and synthetic and natural materials are used to recreate the structure of bone and parts of the human body. Furthermore, metallic biomaterials are used for orthopedic and dental implants because they have superior resistance and load-bearing materials (Hanawa, 2018). Currently, titanium, Co-Cr alloys and 316L Stainless Steel are used in orthopedic applications. Among these metallic materials, Titanium and its alloys have high corrosion resistance, good load-bearing application and excellent mechanical properties along with biocompatibility (Safi, Hussein, Al Shammari, & Tawfiq, 2019) (Walters, 2019).

In addition to their familiar application as hip and knee prostheses, titanium-based materials are also utilized for trauma plates, dental implants, and bone screws. It has been reported that over 1,000 tonnes of titanium-based materials are implanted in the human body worldwide annually, and this quantity is projected to steadily rise over the next decade (Froes, 2018). Ti has traditionally modified with surface engineering treatment to enhance the properties including modifying the surface treatment methods. Surface treatment include anodization, electrophoretic deposition, chemical vapor deposition, physical vapor deposition, etc. (Jiang et al., 2023).

Among these methods, electrophoretic deposition is a mutual colloidal dispensation method often employed in ceramic coating (Obregón, Amor, & Vázquez, 2019). It offers numerous advantages, including a rapid formation process and minimal constraints on substrate shape. Initially, charged particles are dispersed or suspended within a liquid medium and subsequently directed towards an electrode of opposite charge through the application of an electric field, leading to the creation of a deposited film (Pikalova & Kalinina, 2019). For example, Hoomehr et al. produced composite core-shell nanoparticles of bioactive glass-zirconia and effectively applied them onto a

Ti6Al4V substrate using a single-step EPD process (Hoomehr, Raecissi, Ashrafzadeh, Labbaf, & Kharaziha, 2021)....

An alternative method of metal oxide coating has enhanced antibacterial activity with the composite coating. There are reports of Sn coated on Ti metal. Interestingly Tibayan et al, have studied that tin oxide nanoparticles displayed antibacterial characteristics. They evaluated the effectiveness of Ag/SnO₂ nanocomposites as coating materials with strong antibacterial capabilities (Tibayan Jr et al., 2020). In certain situations, antimicrobial coatings are required to maintain high transparency within the visible spectrum for various applications. Various transparent antimicrobial coatings, such as those based on zinc oxide (ZnO, ZnO:Co, ZnO:Sn ZnO-Y₂O₃ ZnO-CeO₂), have been synthesized and examined (Evstropiev et al., 2019). The main drawback of long-term implant failure is bacterial infection associated, during osteointegration process (Hendrijantini et al., 2023). To overcome this drawback, the present work demonstrates that the Electrophoretic deposition (EPD) of SnO/ZnO composite coating on titanium to enhance the corrosion resistance and antibacterial activity.

Materials and Methods

2.1 Preparation of Coating

The medical grade of commercially pure titanium (Cp-Ti) was purchased from Ti Anode Chennai, India. The sample dimension was 1.5 cm x 1.5 cm x 2 mm thickness and the sample was grounded in SiC paper from 250 to 1500 grade to remove debris and to make an even surface. After polishing the sample was kept with acetone and double distilled water using ultrasonication to clean the surface. Then the sample was treated with Kroll's reagent for 10 seconds. They were washed with DD water and kept with a desiccator.

2.2 Electrolyte preparation

The tin chloride (SnCl₂.2H₂O) was purchased from (Merck 98.5% purity) and the Zinc acetate dihydrate (CH₃COO)₂Zn.2H₂O) was purchased from Sigma Aldrich (98 % purity). The 0.1 M of NaOH was prepared for 50 ml DD water and 0.3 g of (CTAB) was added to the solution. Then 0.5 g of tin chloride was added with a mass ratio of 1:1 of zinc acetate solution after the solution was stirred in 1 h. The solution was transferred to an autoclave for the hydrothermal process at 10 h at 90° C. Then the solution was examined by the EPD process. In the EPD process, the potential was applied at 20 V for 10 min, where platinum was the cathode and Ti was the anode to deposited coating. Further, the sample was used in characterization studies

2.3 Surface Characterization studies

The coated sample was examined by the surface topography analysis using Field Emission-Scanning Electron Microscopy with Energy Dispersive X-ray analysis (FE-SEM/EDX) and the JEOL model (JSM-IT800 NANO SEM). The presence of the functional group at the coated sample was confirmed by the FT-IR spectrum using an Alpha II Bruker model spectrometer from the wave number of 4000-400 cm⁻¹ range.

2.4 Hemocompatibility

According to the established procedure outlined by [11], a hemolysis test was conducted. To do this, trisodium citrate (3.2%) was mixed with human blood collected from donors in the correct 9:1 ratio. The samples were then incubated with phosphate buffer saline (PBS) for 30 minutes at 37 °C. Subsequently, anti-coagulated blood was loaded into standard tubes and incubated at 37 °C. After incubation, the tubes were centrifuged at 3000 rpm for 10 minutes, using a positive control of 0.1% sodium carbonate and anti-coagulated blood, as well as a negative control with PBS. The following calculations were performed to determine the hemolysis rate.

2.5 Antibacterial activity

Two types of bacteria the Gram-negative strain *E. coli* and the Gram-positive strain *S. aureus* were investigated in this study. These bacterial cultures were retrieved from frozen stock and then transferred to Trypcase Soy Agar (TSA) plates. The plates were subsequently placed in an incubator at 37 °C for a duration of 18–24 hours. Following this incubation period, the bacteria were transferred to 50 mL of sterile Tryptic Soy Broth (TSB) medium and allowed to grow for 18–24 hours at 37 °C with agitation at 80 rpm. Before inoculation, the bacterial strains were subcultured into fresh TSB at a 1:50 ratio and incubated for 2 hours at 37 °C with agitation at 80 rpm.

Results

3.1 Surface Morphology analysis

The surface morphology and elemental analysis of the coated sample are given in Fig.1. The surface exhibits a laminar array with petal-like morphology on the coated SnO/ZnO on Cp-Ti surface. The elemental dispersive analysis confirms that the Sn, Zn, O, Cl and Na revealed the EDX profile. The surface are interlinked with each other and self-assembly porous structure was formed and the coating shows good adhesion on Cp-Ti surface. Surface morphology plays a role in the osteointegration process. The FE-SEM reveals porous morphology under high magnification, which aid to form the osteointegration process.

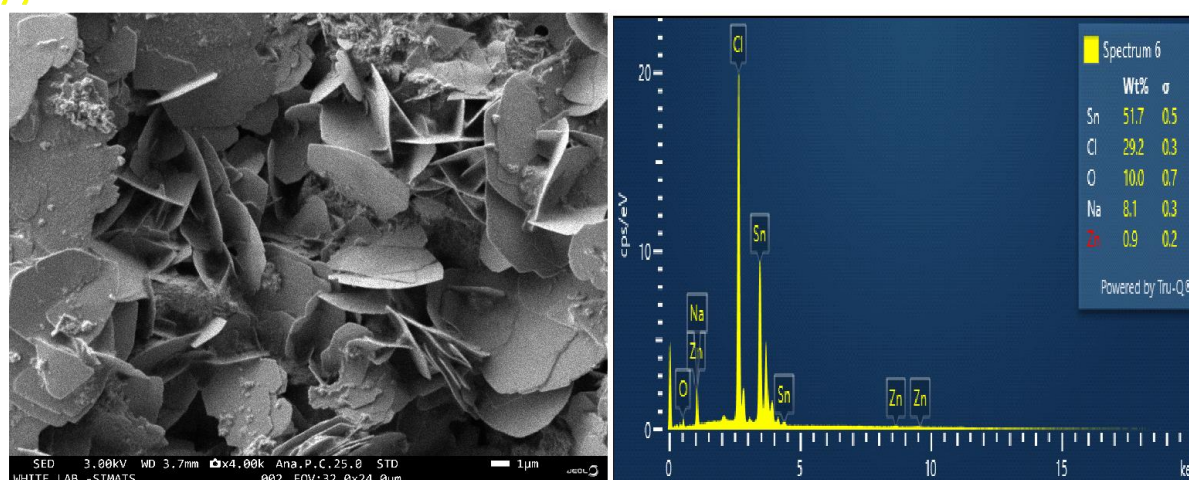


Figure 1. EPD coating of a) SEM and b) EDX profile of SnO/ZnO coated Ti

3.2 ATR-IR studies

In order to identify the functional group analysis of the coated sample was examined by FT-IR spectra in Fig.2. The stretching vibration of the hydroxyl group (OH) and water molecules appeared at the broadband in the wavenumber of $3000-3500\text{ cm}^{-1}$ and 1610 cm^{-1} respectively. The peak ascribed around 600 to 500 cm^{-1} stretching vibration of SnO and an O-Sn-O bending vibration band appeared (Samsonenko, Zakutevskyy, Khalameida, Charmas, & Skubiszewska-Zięba, 2019). The nanoparticles of the SnO were formed at the peak of 1325 cm^{-1} which was in good agreement with SEM images that form the nanostructure (Suresh et al., 2020). A small intensity peak was observed at the ZnO indicating the 597 cm^{-1} (Alamdari et al., 2020). The base metal oxide of the (Ti-O) peak overlapped with the ZnO. This spectrum was confirmed the SnO/ZnO-coated Cp-Ti metal.

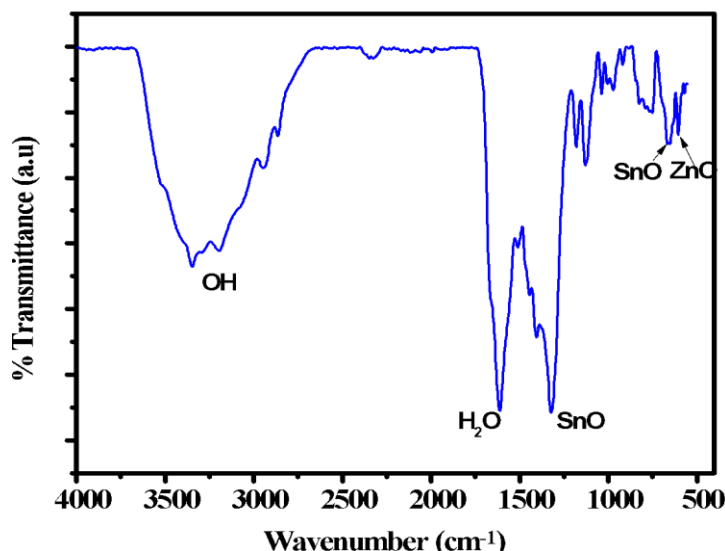


Figure 2. FT-IR spectra of SnO/ZnO coated Cp-Ti

3.3 Hemolysis

The detrimental impact of biomaterial toxicity directly influences Red Blood Cells (RBCs), resulting in hemolysis. When the bone implant material is initially implanted, it interfaces with the blood tissues during the coagulation process. In our study, the hemolysis of the uncoated (Ti) sample and SnO/ZnO coated Ti sample evaluated are displayed in Fig. 3. Further evaluation of the uncoated (Ti) sample exhibits the hemolytic nature of the 7.8 % lysis rate. EPD coating of SnO/ZnO on the Ti surface revealed only hemolytic nature 3.8 % lysis rate. According to the ASTM standard hemocompatibility $< 2\%$ is non-hemolytic, $< 2-5\%$ is slightly hemolytic and above 5% is hemolytic nature (Kalaiyaran, Pugalmani, & Rajendran, 2023). In our study the material showed slightly hemolytic nature, and thus confirming it as biocompatible material which can be used for implant applications.

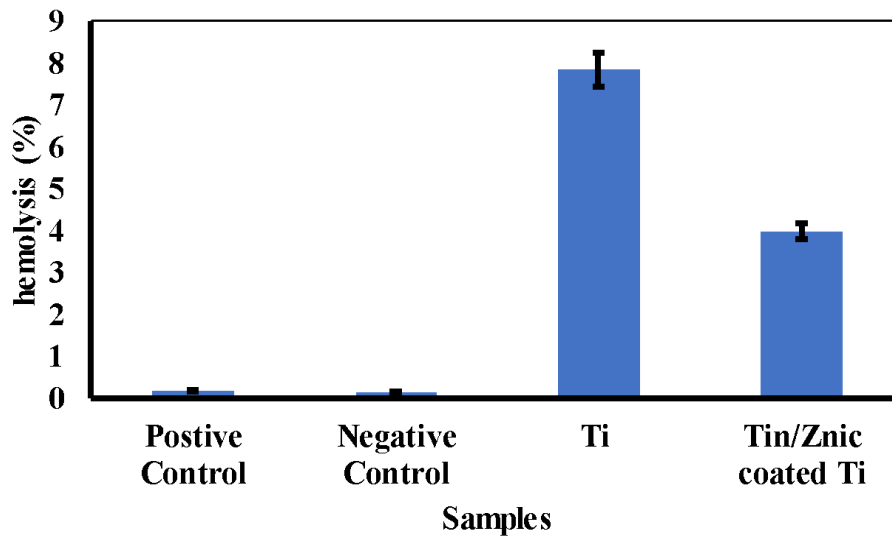


Figure 3. Hemocompatibility of uncoated and coated Ti samples

3.4 Anti-bacterial activity

The antibacterial activity of the coated sample was evaluated by the Zone of inhibition (ZOI) method. Initially, the samples were immersed in PBS solution for 24 h incubation, the extract was used as different concentrations were evaluated and the control used as an antibiotic in Fig.4. The zone of inhibition percentage observed at 25 μ L exhibited the *S. aureus* and *E. coli* exhibits 11 and 10 % respectively. The 50 μ L concentration revealed that 13 and 15 % of the zone. The bacteriostatic force of SnO/ZnO particles killed the cell membrane of bacteria resulting in cell death (Murali et al., 2021).

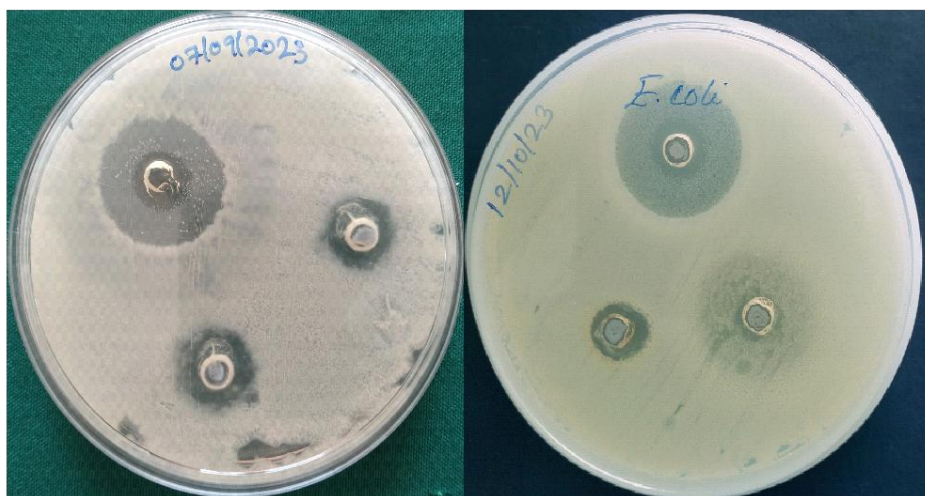
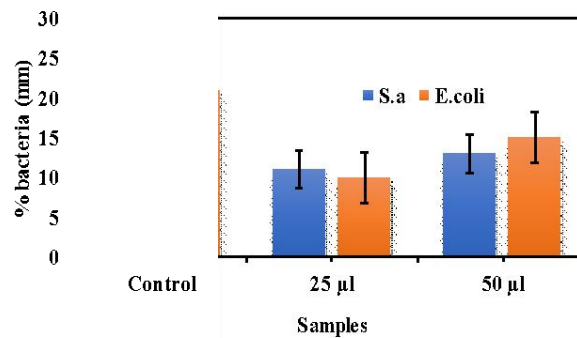


Figure 4. a) Percentage of the zone and b) Zone of the inhibition plate method of *S.aureus* and *E.coli* of coated Ti sample

Discussion

The antibacterial properties of the coating on Cp-Ti are very important for clinical applications. Nowadays, titanium is used the most common implant for orthopedic and dental implants. These implants may fail due to the antibacterial and biofilm formation of the initial stage, before the osteointegration process. In our study coating, the electrophoretic deposition of the Ti sample was coated on SnO/ZnO for applying a potential of 20 V for 10 min. A Bloniarz *et al.*, have fabricated SnO/Sr with bioglass on Ti substrate to enhance the antibacterial activity and bioactivity of the coating (Bloniarz, Cholewa-Kowalska, Gajewska, Grysakowski, & Moskalewicz, 2022). The deposition of the surface was analyzed by the laminar layer and a porous patel-like structure was obtained. The functional group was confirmed by the presence of SnO/ZnO.

The β type of Ti alloy has demonstrated the TiO₂ nanotubes for electrochemical processes and incorporation of Sn reduces the bacterial attachment used in orthopedic applications (Verissimo, Geilich, Oliveira, Caram, & Webster, 2015). In their study, Henry *et al.* investigated the antibacterial properties of SnO₂ thin films were applied to a glass substrate using the sol-gel spin coating technique. They showed antibacterial property of thin SnO₂ films against *Escherichia coli* and Bacillus, as assessed by the agar method. Bacterial growth was observed on the uncoated substrate, but no bacterial growth was detected on the SnO₂-coated substrate. These indicate that SnO₂ offers a promising solution for combatting bacteria through electrostatic interactions that disrupt bacterial membrane integrity and stimulate the production of harmful free radicals (Henry, Mohanraj, Sivakumar, & Umamaheswari, 2015).

The hemocompatibility of titanium and its alloy has outstanding properties required for blood contact implants caused by thrombosis and restenosis. The SnO/ZnO coating has been proved to be the blood-compatible coating. (Manivasagam, Sabino, Kantam, & Popat, 2021). Georgeta *et al.*, have studied the Co-doped ZnO to enhance the antibacterial and in-vitro bioactivity (Voicu *et al.*, 2020). We observe also similar antibacterial activity of Ti-coated samples.

Conclusion

In this study, electrophoretic deposition of SnO/ZnO coated on the Cp-Ti is a promising biocompatible coating which has proven to enhance the surface properties and antibacterial activity of the implant.

Acknowledgment

The authors thank the Instrumentation facilities provided by Saveetha Dental college and Hospitals, Saveetha Institute of Medical and Technical sciences, Chennai, India are gratefully acknowledged.

Conflicts of interest

The authors have no conflicts of interest.

REFERENCES

1. Alamdari, S., Sasani Ghamsari, M., Lee, C., Han, W., Park, H.-H., Tafreshi, M. J, Ara, M. H. M. (2020). Preparation and characterization of zinc oxide nanoparticles using leaf extract of *Sambucus ebulus*. *Appl. Sci.*, 10(10), 3620. <https://doi.org/10.3390/app10103620>
2. Bloniarz, A., Cholewa-Kowalska, K., Gajewska, M., Grysakowski, B., & Moskalewicz, T. (2022). Electrophoretic deposition, microstructure and selected properties of nanocrystalline SnO₂/Sr enriched bioactive glass/chitosan composite coatings on titanium. *Surf. Coat. Technol.*, 450, 129004. <https://doi.org/10.1016/j.surfcoat.2022.129004>
3. Evstropiev, S., Karavaeva, A., Petrova, M., Nikonorov, N., Vasilyev, V., Lesnykh, L., & Dukelskii, K. (2019). Antibacterial effect of nanostructured ZnO-SnO₂ coatings: The role of microstructure. *Mater. Today Commun.*, 21, 100628. <https://doi.org/10.1016/j.mtcomm.2019.100628>
4. Froes, F. S. (2018). Titanium for medical and dental applications—An introduction Titanium in medical and dental applications (pp. 3-21): Elsevier. <https://doi.org/10.1016/B978-0-12-812456-7.00001-9>
5. Hanawa, T. (2018). Transition of surface modification of titanium for medical and dental use Titanium in Medical and Dental Applications (pp. 95-113): Elsevier. <https://doi.org/10.1016/B978-0-12-812456-7.00005-6>
6. Hendrijantini, N., Kuntjoro, M., Agustono, B., Sitalaksmi, R. M., Ari, M. D. A., Theodora, M., . . . Sosiawan, A. (2023). Human umbilical cord mesenchymal stem cells induction in peri-implantitis *Rattus norvegicus* accelerates and enhances osteogenesis activity and implant osseointegration. *Saudi Dent J.*, 35(2), 147-153. <https://doi.org/10.1016/j.sdentj.2023.01.003>
7. Henry, J., Mohanraj, K., Sivakumar, G., & Umamaheswari, S. (2015). Electrochemical and fluorescence properties of SnO₂ thin films and its antibacterial activity. *Spectrochim. Acta A*, 143, 172-178. <https://doi.org/10.1016/j.saa.2015.02.034>
8. Hoomehr, B., Raeissi, K., Ashrafizadeh, F., Labbaf, S., & Kharaziha, M. (2021). Electrophoretic deposition of bioactive glass/zirconia core-shell nanoparticles on Ti6Al4V substrate. *Ceram. Int.*, 47(24), 34959-34969. <https://doi.org/10.1016/j.ceramint.2021.09.037>
9. Jiang, P., Zhang, Y., Hu, R., Shi, B., Zhang, L., Huang, Q., Yang, Y., Tang, P, Lin., C. (2023). Advanced surface engineering of titanium materials for biomedical applications: From static modification to dynamic

- responsive regulation. *Bioact. Mater.*, 27, 15-57. <https://doi.org/10.1016/j.bioactmat.2023.03.006>
10. Kalaiyarasan, M., Pugalmani, S., & Rajendran, N. (2023). Fabrication of chitosan/silica hybrid coating on AZ31 Mg alloy for orthopaedic applications, *J. Magnes. Alloy.*, 11(2), 614-628. <https://doi.org/10.1016/j.jma.2022.05.003>
 11. Manivasagam, V. K., Sabino, R. M., Kantam, P., & Papat, K. C. (2021). Surface modification strategies to improve titanium hemocompatibility: A comprehensive review. *Mater. Adv.*, 2(18), 5824-5842. DOI: 10.1039/D1MA00367D
 12. Murali, M., Kalegowda, N., Gowtham, H. G., Ansari, M. A., Alomary, M. N., Alghamdi, S., N .Shilpa., S.B. Singh., M.C.Triveni., M. Aiyaz., N. Angaswany., N. Lakshmidivi., S.F Adil., M.R. Hatshan., K.M. Amruthgesh., Aiyaz, M. (2021). Plant-mediated zinc oxide nanoparticles: Advances in the new millennium towards understanding their therapeutic role in biomedical applications. *Pharmaceutics*, 13(10), 1662. <https://doi.org/10.3390/pharmaceutics13101662>.
 13. Obregón, S., Amor, G., & Vázquez, A. (2019). Electrophoretic deposition of photocatalytic materials. *Adv. Colloid Interface Sci.*, 269, 236-255. <https://doi.org/10.1016/j.cis.2019.05.003>
 14. Pikalova, E. Y., & Kalinina, E. (2019). Electrophoretic deposition in the solid oxide fuel cell technology: Fundamentals and recent advances. *Renew. Sustain. Energy Rev.*, 116, 109440. <https://doi.org/10.1016/j.rser.2019.109440>
 15. Safi, I. N., Hussein, B. M. A., Al Shammari, A. M., & Tawfiq, T. A. (2019). Implementation and characterization of coating pure titanium dental implant with sintered β -TCP by using Nd: YAG laser. *Saudi Dent J*, 31(2), 242-250. <https://doi.org/10.1016/j.sdentj.2018.12.004>
 16. Samsonenko, M., Zakutevskyy, O., Khalameida, S., Charnas, B., & Skubiszewska-Zięba, J. (2019). Influence of mechanochemical and microwave modification on ion-exchange properties of tin dioxide with respect to uranyl ions. *Adsorption*, 25, 451-457. <https://doi.org/10.1007/s10450-019-00036-2>
 17. Suresh, K., Surendhiran, S., Manoj Kumar, P., Ranjth Kumar, E., Khadar, Y. S., & Balamurugan, A. (2020). Green synthesis of SnO₂ nanoparticles using *Delonix elata* leaf extract: Evaluation of its structural, optical, morphological and photocatalytic properties. *Applied Sciences*, 2, 1-13. <https://doi.org/10.1007/s42452-020-03534-z>
 18. Tibayan Jr, E. B., Muflikhun, M. A., Kumar, V., Fisher, C., Al Rey, C. V., & Santos, G. N. C. (2020). Performance evaluation of Ag/SnO₂ nanocomposite materials as coating material with high capability on antibacterial activity. *Ain Shams Eng. J.*, 11(3), 767-776. <https://doi.org/10.1016/j.asej.2019.11.009>
 19. Verissimo, N. C., Geilich, B. M., Oliveira, H. G., Caram, R., & Webster, T. J. (2015). Reducing *Staphylococcus aureus* growth on Ti alloy nanostructured surfaces through the addition of Sn. *J Biomed Mater Res A.*, 103(12), 3757-3763. <https://doi.org/10.1002/jbm.a.35517>
 20. Voicu, G., Miu, D., Ghitulica, C.-D., Jinga, S.-I., Nicoara, A.-I., Busuioc, C., & Holban, A.-M. (2020). Co doped ZnO thin films deposited by spin coating as antibacterial coating for metallic implants. *Ceram. Int.*, 46(3), 3904-3911. <https://doi.org/10.1016/j.ceramint.2019.10.118>
 21. Walters, N. A. (2019). *Lubrication of Intermetallic Nickel Titanium Bearing Materials*: University of California, Merced.
 22. Williams, D. F. (2009). On the nature of biomaterials. *Biomaterials*, 30(30), 5897-5909.